Amendments to the Claims:

This listing of claims will replace all prior versions, and listings, of claims in the application:

Listing of Claims:

1. (Previously Amended) A method for in vitro regeneration of poinsettia plants comprising:
   (a) incubating poinsettia plant tissue explants that produce reddish epidermal callus on auxin- and cytokinin-containing callus induction medium;
   (b) subculturing reddish epidermal callus to embryo induction medium comprising casein hydrolysate and further comprising NH_4^+ and/or NO_3^- to form embryogenic callus;
   (c) culturing said embryogenic callus on developmental medium containing an osmotic pressure increasing agent and cytokinin;
   (d) culturing said embryogenic callus on maturation medium comprising abscisic acid; and
   (e) recovering poinsettia plants from said embryos.

2. (Original) The method of claim 1, wherein said callus induction medium comprises about 0.5 - 0.8 mg/liter 1-naphthalene acetic acid, about 0.2 - 0.4 mg/liter 6-benzylaminopurine 400 to 1700mb/liter NH_4NO_3, 1900 to 3500 mg/liter KNO_3 and about 1 gm liter casein hydrosylate.

3. (Original) The method of claim 1, wherein said embryo induction medium comprises about 0.5 - 0.8 mg/liter 1-naphthalene acetic acid, about 0.2 - 0.4 mg/liter 6-benzylaminopurine 400 to 1700mb/liter NH_4NO_3, 1900 to 3500 mg/liter KNO_3 and about 1 gm liter casein hydrosylate.

4. (Original) The method of claim 1, wherein said developmental medium comprises about 0.05 mg/liter 6-benzylaminopurine, and about 10 gm/liter mannitol.

5. (Original) The method of claim 1, wherein said maturation medium comprises about 5-20 µM abscisic acid, about 30-100gm/liter sucrose, about 1 gm/liter casein hydrosylate, and about 10 gm/liter mannitol.

6. (Currently Amended) A method for producing transgenic poinsettia plants, comprising:
   (a) incubating poinsettia plant tissue explants that produce reddish epidermal callus on auxin- and cytokinin-containing callus induction medium;

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(b) culturing reddish epidermal callus on embryo induction medium comprising casein hydrolysate and further comprising NH_4^+ and/or NO_3 to form embryogenic callus;

(c)

(i) introducing an expression vector into said incubating embryogenic callus to produce transformed embryogenic callus, wherein said expression vector comprises a selectable marker gene and a second foreign gene, or

(ii) introducing two expression vectors into said incubating embryogenic callus to produce transformed embryogenic callus, wherein one of said expression vectors comprises a selectable marker gene, and wherein the second of said expression vectors comprises a second foreign gene;

wherein the vector or vectors of (c)(i) and (c)(ii) are introduced into the incubating embryogenic callus ~~by co-incubating the callus with Agrobacterium tumefaciens containing the vector or vectors or~~ by microprojectile-mediated delivery of the vector or vectors into the callus;

(d) culturing said transformed embryogenic callus on selection medium;

(e) culturing said transformed embryogenic callus containing embryos on developmental medium containing an osmotic pressure increasing agent;

(f) culturing said transgenic embryos on maturation medium; and

(g) recovering transgenic plants from said transgenic embryos.

7. (Original) The method of claim 6, wherein said callus induction medium comprises about 0.5 - 0.8 mg/liter 1-naphthalene acetic acid, about 0.2 - 0.4 mg/liter 6-benzylaminopurine and about 1 gm liter casein hydrosylate.

8. (Original) The method of claim 6, wherein said embryo induction medium comprises about 0.5 - 0.8 mg/liter 1-naphthalene acetic acid, about 0.2 - 0.4 mg/liter 6-benzylaminopurine and about 1 gm liter casein hydrosylate.

9. (Original) The method of claim 6, wherein said developmental medium comprises about 0.05 mg/liter 6-benzylaminopurine, and about 10 gm/liter mannitol.

10. (Original) The method of claim 6, wherein said maturation medium comprises about 5-20 μM abscisic acid, about 30-100gm/liter sucrose, about 1 gm/liter casein hydrosylate, and about 10 gm/liter mannitol.

11. (Original) The method of claim 8 wherein said embryo induction medium further comprises about 400 to 1700mg/liter NH_4NO_3, 1900 to 3500 mg/liter KNO_3.

12. (Original) The method of claim 6, wherein said poinsettia plant tissue explants are selected from the group consisting of immature embryos, mature embryos, shoot tips and stem segments.

13. (Original) The method of claim 6, wherein said selectable marker gene is selected from the group consisting of a neomycin phosphotransferase gene, a hygromycin phosphotransferase gene, a phosphinothricin gene, a dihydrofolate reductase gene, a 5-enolpyruvylshikimate-3-phosphate synthase gene, an acetohydroxyacid synthase gene, a chloramphenicol acetyltransferase gene, a 3"-adenylyltransferase gene, a gentamicin acetyltransferase gene, a streptomycin phosphotransferase gene, and an aminoglycoside-3'-adenyl transferase gene.

14. (Original) The method of claim 13, wherein said selectable marker gene in hygromycin phosphotransferase and said selection agent is hygromycin.

15. (Original) The method of claim 6, wherein said expression vector that comprises said second foreign gene further comprises a promoter, wherein said promoter is selected from the group consisting of Cauliflower Mosaic Virus (CaMV) 35S promoter, the enhanced 35S promoter, the UBQ3 promoter, the UBQ10 promoter, the UBQ11 promoter, the UBQ14 promoter, the TEFA 1 promoter, the rolC promoter, and the Commelina Yellow Mottle Virus promoter, wherein the expression of said second foreign gene is under the control of said promoter.

16. (Original) The method of claim 15, wherein said promoter is selected from the group consisting of the CaMV 35S promoter, the enhanced 35S promoter, the UBQ3 promoter, and the UBQ10 promoter.

17. (Previously Amended) The method of claim 6, wherein the expression of said second foreign gene confers resistance to disease caused by an organism selected from the group consisting of virus, bacterium, and fungus.

18. (Previously Amended) The method of claim 17, wherein said second foreign gene disrupts the function of said virus, and wherein said virus-disrupting gene is selected from the group consisting of genes encoding viral coat protein, 2'-5' oligonucleotide synthetase, viral genome antisense RNA, and pokeweed antiviral protein.

19. (Previously Amended) The method of claim 6, wherein said second foreign gene confers resistance to an insect, and wherein said insect resistance gene encodes a protein selected from the group consisting of tryptophan decarboxylase, lectin, and Bacillus thuringiensis toxin.

20. (Original) The method of claim 19 wherein said lectin is Galanthus nivalis lectin.

21. (Original) The method of claim 6, wherein said second foreign gene confers resistance to a bacterium or a fungus, and wherein said second foreign gene encodes a polypeptide selected from the group consisting of chitinase, a β-1,3-glucanase, ribosome-inactivating protein, lytic peptide, and plant defensin.

22. (Original) The method of claim 21, wherein said plant defensin is radish seed Rs-AFP2.

23. (Original) The method of claim 21, wherein said lytic peptide is selected from the group consisting of a magainin, PGLa, PGL, xenopsin, caerulein, cecropin, MSI-99, MSI-55, and D5-C.

24. (Original) The method of claim 6, wherein said second foreign gene is operatively linked with a DNA molecule encoding pea vicilin signal peptide.

25. (Original) The method of claim 23, wherein said magainin is magainin 1 or magainin 2.

26. (Original) The method of claim 6, wherein said transgenic poinsettia comprises an expression vector that further comprises a third foreign gene.

27. (Original) The method of claim 6, wherein said second foreign gene encodes chitinase, and wherein said third foreign gene encodes β-1,3-glucanase.

28. (Original) The method of claim 6, wherein said second foreign gene encodes magainin 2, and wherein said third foreign gene encodes PGLa or PGL.

29. (Original) The method of claim 6, wherein the expression of said second foreign gene confers insensitivity to ethylene, and wherein said second foreign gene encodes a mutated ethylene receptor.

30. (Original) The method of claim 29, wherein said mutated ethylene receptor gene is the Arabidopsis etr-1 gene or a tomato NR gene.

31. (Original) The method of claim 6, wherein said second foreign gene is the Vitreoscilla hemoglobin gene.

32. (Original) The method of claim 6, wherein said second foreign gene is an isopentyenyl transferase gene, wherein the expression of said isopentynyl transferase gene is under the control of a promoter of a senescence-associated gene.

33. (Original) The method of claim 32, wherein said promoter is the Arabidopsis SAG12 gene promoter.

34. (Original) The method of claim 6, wherein said second foreign gene encodes a polypeptide having a MADS box domain.

35. (Original) The method of claim 34, wherein said second foreign gene is selected from the group consisting of the PLENA gene, the SQUAMOSA gene, the DEFICIENS A gene, the GLOBOSA gene, the APTELA1 gene, the APETALA2 gene, the AGAMOUS gene, the OsMADS24 gene, the OsMADS45 gene, and the OsMADS1 gene.

36. (Original) The method of claim 6, wherein said foreign gene encodes a protein that modifies plant habit.

37. (Original) The method of claim 36, wherein said gene is the OsMADS1 or phyA gene.

38. (Canceled) The method of claim 6, wherein said expression vector is introduced by microparticle bombardment.

39. (Currently Amended) A method for producing transgenic poinsettia plants, comprising:

(a) incubating poinsettia plant tissue explants that produce reddish epidermal callus in auxin- and cytokinin-containing callus induction medium;

(b) subculturing embryogenic callus produced on said callus induction medium to liquid NH_4^+ and/or NO_3^- containing embryo induction medium comprising casein hydrolysate;

(c) filtering the culture and culturing the filtrate in fresh liquid embryo induction medium;

(d) filtering the culture and culturing the filtrate on solid embryo induction medium;

(e) subculturing embryos produced on said embryo induction medium to maturation medium;

(f) culturing said embryos on callus induction medium;

(g) subculturing epidermal callus produced on said callus induction medium to embryo induction medium to form embryogenic callus;

(h)

(i) introducing an expression vector into said embryogenic callus to produce transformed embryogenic callus, wherein said expression vector comprises a selectable marker gene and a second foreign gene, or

(ii) introducing two expression vectors into said embryogenic callus to produce transformed embryogenic callus, wherein one of said expression vectors comprises a selectable marker gene, and wherein the second of said expression vectors comprises a second foreign gene;

wherein the vector or vectors of (h)(i) and (h)(ii) are introduced into the incubating embryogenic callus ~~by co-incubating the callus with Agrobacterium tumefaciens containing the vector or vectors or~~ by microprojectile-mediated delivery of the vector or vectors into the callus;

(i) culturing said transformed embryogenic callus on selection medium;

(j) culturing said transformed embryogenic callus containing embryos on developmental medium containing an osmotic pressure increasing agent;

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(k) culturing said transformed embryos on maturation medium; and

(l) recovering transgenic plants from said transgenic embryos.

40. (Original) The method of claim 39, wherein said callus induction medium comprises about 0.5 - 0.8 mg/liter 1-naphthalene acetic acid, about 0.2 - 0.4 mg/liter 6-benzylaminopurine and about 1 gm liter casein hydrosylate.

41. (Original) The method of claim 39, wherein said embryo induction medium comprises about 0.5 - 0.8 mg/liter 1-naphthalene acetic acid, about 0.2 - 0.4 mg/liter 6-benzylaminopurine and about 1 gm liter casein hydrosylate.

42. (Original) The method of claim 39, wherein said developmental medium comprises about 0.05 mg/liter 6-benzylaminopurine, and about 10 gm/liter mannitol.

43. (Original) The method of claim 39, wherein said maturation medium comprises about 5-20 μM abscisic acid, about 30-100gm/liter sucrose, about 1 gm/liter casein hydrosylate, and about 10 gm/liter mannitol.

44. (Original) The method of claim 40 wherein said embryo induction medium further comprises about 400 to 1700mg/liter NH_4NO_3, 1900 to 3500 mg/liter KNO_3.

45. (Original) The method of claim 39, wherein said poinsettia plant tissue explants are selected from the group consisting of immature embryos, mature embryos, shoot tips and stem segments.

47. (Original) The method of claim 39, wherein said selectable marker gene is selected from the group consisting of a neomycin phosphotransferase gene, a hygromycin phosphotransferase gene, a phosphinothricin gene, a dihydrofolate reductase gene, a 5-enolpyruvylshikimate-3-phosphate synthase gene, an acetohydroxyacid synthase gene, a chloramphenicol acetyltransferase gene, a 3''-adenylyltransferase gene, a gentamicin acetyltransferase gene, a streptomycin phosphotransferase gene, and an aminoglycoside-3'-adenyl transferase gene.

48. (Original) The method of claim 47, wherein said selectable marker gene is hygromycin phosphotransferase and said selection agent is hygromycin.

49. (Original) The method of claim 47, wherein said expression vector that comprises said second foreign gene further comprises a promoter, wherein said promoter is selected from the group consisting of Cauliflower Mosaic Virus (CaMV) 35S promoter, the enhanced 35S promoter, the UBQ3 promoter, the UBQ10 promoter, the UBQ11 promoter, the UBQ14 promoter, the TEFA 1 promoter, the rolC promoter, and the Commelina Yellow Mottle Virus promoter, wherein the expression of said second foreign gene is under the control of said promoter.

50. (Original) The method of claim 49, wherein said promoter is selected from the group consisting of the CaMV 35S promoter, the enhanced 35S promoter, the UBQ3 promoter, and the UBQ10 promoter.

51. (Previously Amended) The method of claim 39, wherein the expression of said second foreign gene confers resistance to disease caused by an organism selected from the group consisting of virus, bacterium, and fungus.

52. (Previously Amended) The method of claim 51, wherein said second foreign gene disrupts the function of said virus, and wherein said virus-disrupting gene is selected from the group consisting of genes encoding viral coat protein, 2'-5' oligonucleotide synthetase, viral genome antisense RNA, and pokeweed antiviral protein.

53. (Previously Amended) The method of claim 39, wherein said second foreign gene confers resistance to an insect, and wherein said insect resistance gene encodes a protein selected from the group consisting of tryptophan decarboxylase, lectin, and Bacillus thuringiensis toxin.

54. (Original) The method of claim 53 wherein said lectin is Galanthus nivalis lectin.

55. (Original) The method of claim 39, wherein said second foreign gene confers resistance to a bacterium or a fungus, and wherein said second foreign gene encodes a polypeptide selected from the group consisting of chitinase, a β-1,3-glucanase, ribosome-inactivating protein, lytic peptide, and plant defensin.

56. (Original) The method of claim 55, wherein said plant defensin is radish seed Rs-AFP2.

57. (Original) The method of claim 55, wherein said lytic peptide is selected from the group consisting of a magainin, PGLa, PGL, xenopsin, caerulein, cecropin, MSI-99, MSI-55, and D5-C.

58. (Original) The method of claim 39, wherein said second foreign gene is operatively linked with a DNA molecule encoding pea vicilin signal peptide.

59. (Original) The method of claim 39, wherein said magainin is magainin 1 or magainin 2.

60. (Original) The method of claim 39, wherein said transgenic poinsettia comprises an expression vector that further comprises a third foreign gene.

61. (Original) The method of claim 39, wherein said second foreign gene encodes chitinase, and wherein said third foreign gene encodes β-1,3-glucanase.

62. (Original) The method of claim 39, wherein said second foreign gene encodes magainin 2, and wherein said third foreign gene encodes PGLa or PGL.

63. (Original) The method of claim 39, wherein the expression of said second foreign gene confers insensitivity to ethylene, and wherein said second foreign gene encodes a mutated ethylene receptor.

64. (Original) The method of claim 63, wherein said mutated ethylene receptor gene is the Arabidopsis etr-1 gene or a tomato NR gene.

65. (Original) The method of claim 39, wherein said second foreign gene is the Vitreoscilla hemoglobin gene.

66. (Original) The method of claim 39, wherein said second foreign gene is an isopentyenyl transferase gene, wherein the expression of said isopentynyl transferase gene is under the control of a promoter of a senescence-associated gene.

67. (Original) The method of claim 66, wherein said promoter is the Arabidopsis SAG12 gene promoter.

68. (Original) The method of claim 39, wherein said second foreign gene encodes a polypeptide having a MADS box domain.

69. (Original) The method of claim 68, wherein said second foreign gene is selected from the group consisting of the PLENA gene, the SQUAMOSA gene, the DEFICIENS A gene, the GLOBOSA gene, the APTELA1 gene, the APETALA2 gene, the AGAMOUS gene, the OsMADS24 gene, the OsMADS45 gene, and the OsMADS1 gene.

70. (Original) The method of claim 39, wherein said foreign gene encodes a protein that modifies plant habit.

71. (Original) The method of claim 70, wherein said gene is the OsMADS1 or phyA gene.

72. (Canceled) The method of claim 39, wherein said expression vector is introduced by microparticle bombardment.

73. (Canceled) A transgenic poinsettia plant comprising at least one expression vector, wherein said expression vector comprises at least one foreign gene, and wherein said transgenic poinsettia plant expresses said foreign gene.

74. (Canceled) The transgenic poinsettia plant of claim 73, wherein said expression vector further comprises a promoter, wherein said promoter is selected from the group consisting of Cauliflower Mosaic Virus (CaMV) 35S promoter, the enhanced 35S promoter, the UBQ3 promoter, the UBQ10 promoter, the UBQ11 promoter, the UBQ14 promoter, the TEFA 1 promoter, the rolC promoter, and the Commelina Yellow Mottle Virus promoter, wherein the expression of said foreign gene is under the control of said promoter.

75. (Canceled) The transgenic poinsettia plant of claim 74, wherein said promoter is selected from the group consisting of the CaMV 35S promoter, the enhanced 35S promoter, the UBQ3 promoter, and the UBQ10 promoter.

76. (Previously Amended) The transgenic poinsettia plant of claim 73, wherein the expression of said foreign gene confers resistance to disease caused by an organism selected from the group consisting of virus, bacterium, and fungus.

77. (Previously Amended) The transgenic poinsettia plant of claim 76, wherein said foreign gene disrupts the function of said virus, and wherein said virus-disrupting gene is selected from the

group consisting of genes encoding viral coat protein, 2'-5' oligonucleotide synthetase, viral genome antisense RNA, and pokeweed B81antiviral protein.

78. (Previously Amended) The transgenic poinsettia plant of claim 73, wherein said foreign gene confers resistance to an insect, and wherein said insect resistance gene encodes a protein selected from the group consisting of tryptophan decarboxylase, lectin, and Bacillus thuringiensis toxin.

79. (Original) The transgenic poinsettia plant of claim 78 wherein said lectin is Galanthus nivalis lectin.

80. (Previously Amended) The transgenic poinsettia plant of claim 76, wherein said foreign gene confers resistance to a bacterium or a fungus and encodes a polypeptide selected from the group consisting of chitinase, a β-1,3-glucanase, ribosome-inactivating protein, lytic peptide, and plant defensin.

81. (Original) The transgenic poinsettia plant of claim 80, wherein said plant defensin is radish seed Rs-AFP2.

82. (Original) The transgenic poinsettia plant of claim 80, wherein said lytic peptide is selected from the group consisting of a magainin, PGLa, PGL, xenopsin, caerulein, cecropin, MSI-99, MSI-55, and D5-C.

83. (Canceled) The transgenic poinsettia plant of claim 73, wherein said foreign gene is operatively linked with a DNA molecule encoding pea vicilin signal peptide.

84. (Original) The transgenic poinsettia plant of claim 82, wherein said magainin in magainin 1 or magainin 2.

85. (Canceled) The transgenic poinsettia plant of claim 73, wherein said transgenic poinsettia comprises an expression vector that further comprises a second foreign gene.

86. (Currently Amended) The transgenic poinsettia plant ~~of claim 85,~~ comprising at least one expression vector, wherein said expression vector comprises a first ~~wherein said~~ foreign

gene ~~encodes~~ encoding chitinase, and ~~wherein said~~ a second foreign gene ~~encodes~~ encoding β-1,3-glucanase, wherein the foreign genes are expressed.

87. (Original) The transgenic poinsettia plant of claim 86, wherein said foreign gene encodes magainin 2, and wherein said second foreign gene encodes PGLa or PGL.

88. (Original) The transgenic poinsettia plant of claim 86, wherein the expression of said foreign gene confers insensitivity to ethylene, and wherein said foreign gene encodes a mutated ethylene receptor.

89. (Original) The transgenic poinsettia plant of claim 88, wherein said mutated ethylene receptor gene is the Arabidopsis etr-1 gene or a tomato NR gene.

90. (Original) The transgenic poinsettia plant of claim 73, wherein said foreign gene is the Vitreoscilla hemoglobin gene.

91. (Original) The transgenic poinsettia plant of claim 73, wherein said foreign gene is an isopentyenyl transferase gene, wherein the expression of said isopentynyl transferase gene is under the control of a promoter of a senescence-associated gene.

92. (Original) The transgenic poinsettia plant of claim 91, wherein said promoter is the Arabidopsis SAG12 gene promoter.

93. (Original) The transgenic poinsettia plant of claim 73, wherein said foreign gene encodes a polypeptide having a MADS box domain.

94. (Original) The transgenic poinsettia plant of claim 93, wherein said second foreign gene is selected from the group consisting of the PLENA gene, the SQUAMOSA gene, the DEFICIENS A gene, the GLOBOSA gene, the APTELA1 gene, the APETALA2 gene, the AGAMOUS gene, the OsMADS24 gene, the OsMADS45 gene, and the OsMADS1 gene.

95. (Canceled) The transgenic poinsettia plant of claim 73, wherein said foreign gene encodes a protein that modifies plant habit.

96. (Original) The transgenic poinsettia plant of claim 95, wherein said gene is the OsMADS1 or phyA gene.

97. (Previously Added) The method of claim 1, wherein said poinsettia plants of step (e) are fertile.

98. (Previously Added) The method of claim 6, wherein said poinsettia plants of step (g) are fertile.

99. (Previously Added) The method of claim 39, wherein said poinsettia plants of step (l) are fertile.

100. (Canceled) The transgenic poinsettia plant of claim 73, wherein said plant is fertile.

101. (Canceled) A method for in vitro regeneration of poinsettia plants comprising:

(a) incubating poinsettia plant tissue explants that produce epidermal callus on auxin- and cytokinin-containing callus induction medium;

(b) subculturing reddish epidermal callus to NH_4^+ and/or NO_3^- containing embryo induction medium to form embryogenic callus containing embryos;

(c) culturing said embryogenic callus containing embryos on developmental medium containing an osmotic pressure increasing agent and cytokinin;

(d) culturing said embryogenic callus containing embryos on maturation medium comprising abscisic acid; and

(e) recovering poinsettia plants from said embryos.

102. (Currently Amended) A method for producing transgenic poinsettia plants comprising the steps of:

(a) incubating poinsettia plant tissue explants that produce epidermal callus on auxin- and cytokinin-containing callus induction medium;

(b) subculturing embryogenic callus to embryo induction medium comprising casein hydrolysate and further comprising NH_4^+ and/or NO_3^- to form embryogenic callus containing embryos;

(c)

(i) introducing an expression vector into said incubating embryogenic callus to produce transformed embryogenic callus, wherein said expression vector comprises a selectable marker gene and a second foreign gene, or

(ii) introducing two expression vectors into said incubating embryogenic callus to produce transformed embryogenic callus, wherein one of said expression vectors comprises a selectable marker gene, and wherein the second of said expression vectors comprises a second foreign gene;

wherein the vector or vectors of (c)(i) and (c)(ii) are introduced into the incubating embryogenic callus ~~by co-incubating the callus with Agrobacterium tumefaciens containing the vector or vectors or~~ by microprojectile-mediated delivery of the vector or vectors into the callus;

(d) culturing said transformed embryogenic callus on selection medium;

(e) culturing said embryogenic callus containing embryos on developmental medium containing an osmotic pressure increasing agent;

(f) culturing said transgenic embryos on maturation medium; and

(g) recovering transgenic plants from said transgenic embryos.

103. (Previously Amended) A method for producing transgenic poinsettia plants comprising the steps of:

(a) incubating poinsettia plant tissue explants that produce epidermal callus on auxin- and cytokinin-containing callus induction medium;

(b) subculturing embryogenic callus produced on said callus induction medium to liquid embryo induction medium comprising casein hydrolysate and further comprising NH_4^+ and/or NO_3^-;

(c) filtering the culture and culturing the filtrate in fresh liquid embryo induction medium;

(d) filtering the culture and culturing the filtrate on solid embryo induction medium,

(e) subculturing embryos produced on said embryo induction medium to maturation medium;

(f) culturing said embryos on callus induction medium;

(g) subculturing embryogenic callus produced on said callus induction medium to embryo induction medium to form embryogenic callus containing embryos;

(h)

(i) introducing an expression vector into said incubating embryogenic callus to produce transformed embryogenic callus, wherein said expression vector comprises a selectable marker gene and a second foreign gene, or

(ii) introducing two expression vectors into said incubating embryogenic callus to produce transformed embryogenic callus, wherein one of said expression vectors comprises a selectable marker gene, and wherein the second of said expression vectors comprises a second foreign gene;

wherein the vector or vectors of (h)(i) and (h)(ii) are introduced into the incubating embryogenic callus by microprojectile-mediated delivery of the vector or vectors into the callus;

(i) culturing said transformed embryogenic callus on selection medium;

(j) culturing said transformed embryogenic callus containing embryos on developmental medium containing an osmotic pressure increasing agent;

(k) culturing said transformed embryos on maturation medium; and

(l) recovering transgenic plants from said transgenic embryos.

104. (Canceled) The method of claim 101, wherein said developmental medium comprises about 0.05 mg/liter cytokinin.

105. (Previously Added) The method of claim 102, wherein said developmental medium comprises about 0.05 mg/liter cytokinin.

106. (Previously Added) The method of claim 103, wherein said developmental medium comprises about 0.05 mg/liter cytokinin.

108. (Previously Added) The method of claim 102, wherein said developmental medium comprises cytokinin.

109. (Previously Added) The method of claim 103, wherein said developmental medium comprises cytokinin.

110. (Previously Added) The method of claim 6, wherein the expression of said second foreign gene confers resistance to an insect.

111. (Previously Amended) The method of claim 39, wherein the expression of said second foreign gene confers resistance to an insect.

112. (Canceled) The transgenic poinsettia plant of claim 73, wherein the expression of said second foreign gene confers resistance to an insect.

113 – 118 Canceled.